

TITLE OF THE INVENTION

METHOD OF FORMING AN ARTIFICIAL SHIRO OF MATSUTAKE

CROSS-REFERENCE TO RELATED APPLICATIONS

5 This application is based upon and claims the
benefit of priority from the prior Japanese Patent
Application No. 2001-018691, filed January 26, 2001,
the entire contents of which are incorporated herein by
reference.

BACKGROUND OF THE INVENTION

10 1. Field of the Invention

The present invention relates to a method of
forming an artificial Shiro of *Tricholoma Matsutake*
(hereinafter referred to as "Matsutake").

2. Description of the Related Art

15 Generally, as a method for efficiently cultivating
a large amount of mushrooms, a method of proliferating
mycelia of the mushroom in a liquid culture medium is
known. However, in case of Matsutake mushrooms, their
production requires a root of a living tree, because
20 Matsutake is an ectomycorrhizal fungus. An assembly
of hyphae called a "Shiro" is present in the site where
a Matsutake mushroom grows, and therefore induction
of the Shiro formation is an important step toward
increasing the production of Matsutake mushrooms
25 (Makoto Ogawa, *Biology of Matsutake*, pp 333, published
by Tsukiji Shokan).

However, in a natural environment, saprobe

and sugar fungus preferentially proliferate in the nutrient site where a red pine (i.e., a host plant of a Matsutake fungus) grows. Because of this, it is difficult for a Matsutake fungus (a symbiotic fungus) to form its Shiro.

Various studies have been made on artificial cultivation of Matsutake mushrooms (*Tricholoma Matsutake* (S. Ito et Imai) Sing.) from long ago. Nevertheless, it is still impossible to cultivate Matsutake mushrooms artificially.

The results of studies that have been made on the artificial cultivation of Matsutake mushrooms will be described below.

Masui has experimentally showed that Matsutake forms a mycorrhizal association with a root of a red pine (Masui, K. (1927), Mem. Coll. Sci. Kyoto Univ. B (2) 2, 149-279). Tominaga has investigated the optimal conditions for growth of Matsutake hyphae (Tominaga, Y. (1965) Bull. Hiroshima Agri. Coll. 2: 242-246). Ogawa et al. have investigated the formation of a Matsutake fruit body primordium by axenic culture (Ogawa, M & Hamada, M (1975), Trans. Mycol. Soc. Japan 16: 406-415). However, success in the induction of Matsutake Shiro formation has not yet been reported.

To make the present invention, the present inventors have focused on the fact that the Matsutake hyphae grow extremely slowly. They considered that the

slow growth of the hyphae might be one of the factors that hindered advancements of artificial cultivation of Matsutake mushrooms. Up to now, in an attempt to produce Matsutake mushrooms in a large amount, artificial formation of a new Shiro has been tried by inoculating Matsutake hyphae into a host plant-growing forest. However, by such a trial, Matsutake hyphae have not grown to form a Shiro to the extent of practical use. Therefore, it is presently considered difficult to cultivate Matsutake mushrooms artificially.

BRIEF SUMMARY OF THE INVENTION

In view of the aforementioned background, the present inventors have made extensive studies on a method of inducing the formation of an artificial Shiro of Matsutake, which is an important step toward increasing the production of Matsutake mushrooms. Therefore, an object of the present invention is to provide a method capable of rapidly forming an artificial Shiro of Matsutake.

The present inventors found an active principle capable of promoting the growth of Matsutake hyphae. Based on this finding, they established a method of proliferating Matsutake hyphae in a short time, thereby accomplishing the present invention. More specifically, the present invention is achieved by the means described below.

(1) A method of forming an artificial Shiro of Matsutake comprising:

5 culturing Matsutake hyphae in a culture substrate containing a substance capable of controlling the cell membrane permeability of the hyphae as an active principle.

(2) A method of forming an artificial Shiro of Matsutake comprising:

10 culturing Matsutake hyphae in a culture substrate containing a substance capable of enhancing the hydrophilic property of the hyphae as an active principle.

(3) A method of forming an artificial Shiro of Matsutake comprising:

15 culturing Matsutake hyphae in a culture substrate containing a surfactant and/or a natural vegetable oil as an active principle.

(4) A method of forming an artificial Shiro of Matsutake comprising:

20 culturing Matsutake hyphae in a culture substrate containing a fatty acid ester as an active principle.

(5) A method of forming an artificial Shiro of Matsutake, comprising:

25 inducing growth of Matsutake hyphae by aseptically homogenizing a colony of Matsutake hyphae and aseptically culturing the obtained hyphae in a liquid nutrient medium;

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preparing an inoculum of Matsutake hyphae by aseptically replacing the liquid nutrient medium containing the growth-induced Matsutake hyphae with a liquid nutrient medium containing no carbon source; and

5 culturing aseptically the inoculum of the Matsutake hyphae in a culture substrate containing a substance capable of controlling the cell membrane permeability of the hyphae as an active principle.

(6) A method of forming an artificial Shiro of Matsutake, comprising:

10 inducing growth of Matsutake hyphae by aseptically homogenizing a colony of Matsutake hyphae and aseptically culturing the obtained hyphae in a liquid nutrient medium;

15 preparing an inoculum of Matsutake hyphae by aseptically replacing the liquid nutrient medium containing the growth-induced Matsutake hyphae with a liquid nutrient medium containing no carbon source; and

20 culturing aseptically the inoculum of the Matsutake hyphae in a culture substrate containing a substance capable of enhancing the hydrophilic property of the hyphae as an active principle.

(7) A method of forming an artificial Shiro of Matsutake, comprising:

25 inducing growth of Matsutake hyphae by aseptically homogenizing a colony of Matsutake hyphae and aseptically culturing the obtained hyphae in a liquid

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nutrient medium;

preparing an inoculum of Matsutake hyphae by
aseptically replacing the liquid nutrient medium
containing the growth-induced Matsutake hyphae with a
5 liquid nutrient medium containing no carbon source; and
culturing aseptically the inoculum of the
Matsutake hyphae in a culture substrate containing
a surfactant and/or a natural vegetable oil as
an active principle.

10 (8) A method of forming an artificial Shiro of
Matsutake, comprising:

inducing growth of Matsutake hyphae by aseptically
homogenizing a colony of Matsutake hyphae and
aseptically culturing the obtained hyphae in a liquid
15 nutrient medium;

preparing an inoculum of Matsutake hyphae by
aseptically replacing the liquid nutrient medium
containing the growth-induced Matsutake hyphae with a
liquid nutrient medium containing no carbon source; and
20 culturing aseptically the inoculum of the
Matsutake hyphae in a culture substrate containing
a fatty acid ester as an active principle.

(9) The method of forming an artificial Shiro of
Matsutake according to any one of (1) to (8), wherein
25 a solution containing the active principle at the
concentration of 0.2 to 10 wt% is used as the active
principle.

(10) The method of forming an artificial Shiro of Matsutake according to any one of (1) to (9), wherein a solution containing the active principle which is prepared using an organic solvent and distilled water is used as the active principle.

(11) The method of forming an artificial Shiro of Matsutake according to any one of (1) to (10), wherein either one of soil having a grain size of 3 mm or less and an artificial substrate having a grain size of 2 mm or less is used as the culture substrate.

(12) The method of forming an artificial Shiro of Matsutake according to any one of (1) to (11), wherein the active principle is added to the culture substrate in a state of a solution containing the active principle, and weight ratio of the solution containing the active principle to the total weight is 15 to 30 wt%.

Additional objects and advantages of the invention will be set forth in the description which follows, and in part will be obvious from the description, or may be learned by practice of the invention. The objects and advantages of the invention may be realized and obtained by means of the instrumentalities and combinations particularly pointed out hereinafter.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWING

The accompanying drawings, which are incorporated in and constitute a part of the specification,

illustrate embodiments of the invention, and together with the general description given above and the detailed description of the embodiments given below, serve to explain the principles of the invention.

5 FIG. 1 is a graph showing an increase of the amount of Matsutake hyphae by the addition of Tween 80.

FIG. 2 is a graph showing an increase of the amount of Matsutake hyphae by the addition of olive oil.

10 DETAILED DESCRIPTION OF THE INVENTION

The method of forming an artificial Shiro of Matsutake of the present invention will be explained in detail below.

15 As a Matsutake strain to be used in the present invention, a commercially available strain from ATCC (American Type Culture Collection), such as ATCC 34979, ATCC 34981, and ATCC 34988 may be used. Preferably, a good strain in growth is screened from commercially available strains and it may be put to use.

20 The Matsutake strain can be proliferated in a medium generally used for maintaining a symbiotic fungus in the art, such as Ohta medium (Ohta, A. (1990) Trans. Mycol. Soc. Japan 31: 323-334), MMN medium (Marx, D. H. (1969) Phytopathology 59: 153-163), and
25 Hamada medium (Hamada (1964) Matsutake, 97-100).

Of them, Ohta medium is preferable for proliferating the Matsutake strain from the viewpoint of its

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Several types of such a substance may be used in combination. Such a substance capable of controlling the cell membrane permeability of the hyphae is considered to facilitate release of an enzyme such as cellulase (that is present within the hyphae) from the hyphae. Preferable examples of the substance capable of controlling the cell membrane permeability of the hyphae may be a surfactant and a natural vegetable oil.

In another aspect of the present invention, Matsutake hyphae is cultured in a culture substrate containing a substance capable of enhancing the hydrophilic property of the hyphae as an active principle. In the present invention, the substance capable of enhancing the hydrophilic property of the hyphae is not particularly limited, as long as it promotes intrusion of the hyphae into the culture substrate and contributes to an increase in absorption amount of nutrient by the hyphae. Several types of such a substance may be used in combination. Preferable examples of the substance capable of enhancing the hydrophilic property of the hyphae may be a surfactant and a natural vegetable oil.

In another aspect of the present invention, Matsutake hyphae are cultured in a culture substrate containing a surfactant and/or a natural vegetable oil as an active principle. Either the surfactant or the natural vegetable oil may be contained as an active principle, or alternatively, the both may be contained. In the present invention, as long as a substance falls under the category of the surfactant or the natural vegetable oil, it can be used as the active principle of the present invention even if it does not necessarily control the cell membrane permeability of the hyphae. Similarly, in the present invention, as long as a substance falls under the category of the

surfactant or the natural vegetable oil, it can be used as the active principle of the present invention even if it does not necessarily enhance the hydrophilic property of the hyphae.

5 The surfactant to be used in the present invention is not particularly limited, as long as it is called as a surfactant. Preferably a nonionic surfactant may be used, and more preferably a Tween series surfactant such as Tween 80 and Tween 40 may be used. Several
10 types of surfactants may be used in combination.

 The natural vegetable oil to be used in the present invention is not particularly limited, as long as it is called as a natural vegetable oil. For example, olive oil, safflower oil, or grape-seed
15 oil may be used. Several types of natural vegetable oils may be used in combination.

 The active principle preferably used in the present invention may be also expressed as fatty acid esters, according to another concept. In one aspect
20 of the present invention, Matsutake hyphae are cultured in a culture substrate containing a fatty acid ester as an active principle. In the present invention, the fatty acid ester is preferably a higher fatty acid ester, more preferably a fatty acid ester having a
25 fatty acid of 12-18 carbon atoms, further preferably, an unsaturated fatty acid ester having a fatty acid of 12-18 carbon atoms, and still preferably, oleic acid

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ester. For example, a fat and oil containing the
aforementioned preferable fatty acid may be used as the
active principle. Several types of fatty acid esters
may be used in combination as the active principle.

5 Also, the fatty acid ester may be used in combination
with a substance other than the fatty acid ester,
the substance being capable of controlling the cell
membrane permeability of the hyphae. The fatty acid
ester may be used in combination with a substance other
10 than the fatty acid ester, the substance being capable
of enhancing the hydrophilic property of the hyphae.

The fact that the fatty acid ester can be used
as the active principle in the present invention is
considered to be closely related to the facts that
15 the aforementioned fatty acid ester is contained in
some of the surfactants and the natural vegetable oils.
Therefore, if a substance falls under the category of
the fatty acid ester and the substance is contained in
the surfactant or the natural vegetable oil, naturally
20 it can be used as the active principle in the present
invention.

The aforementioned active principle is preferably
added to a culture substrate in a state of a solution
containing the active principle. The solution
25 containing the active principle such as the substance
capable of controlling the cell membrane permeability
of the hyphae (e.g., a surfactant and/or a natural

vegetable oil) is preferably prepared by diluting it with an organic solvent (preferably ethanol) and distilled water to a desired concentration before being added to a culture substrate. A solution containing the substance capable of controlling the cell membrane permeability of the hyphae is preferably prepared by diluting it to a concentration of 0.2 to 10 wt%. Similarly, a solution containing the substance capable of enhancing the hydrophilic property of the hyphae is preferably prepared by diluting it to a concentration of 0.2 to 10 wt%.

More specifically, when a surfactant is used as the active principle, the dilution concentration of the surfactant is generally 0.2 wt% or more, preferably 0.2 to 10 wt%, and more preferably 1 to 5 wt%. The dilution concentration of less than 0.2 wt% is not preferable for the reason that the remarkable effect of the surfactant on growth of the hyphae cannot be brought about.

When a natural vegetable oil is used as the active principle, the dilution concentration of the natural vegetable oil is generally 0.2 wt% or more, preferably 0.2 to 5 wt%, and more preferably 0.5 to 2 wt%. Similarly, the dilution concentration of less than 0.2 wt% is not preferable for the reason that the remarkable effect of the natural vegetable oil on growth of the hyphae cannot be brought about.

In the case where the surfactant and the natural vegetable oil are used together as the active principle, the dilution concentration is set at generally 0.2 wt% or more, preferably 0.2 to 10 wt%, and more preferably 0.5 to 5 wt%, in total.

When a fatty acid ester is used as the active principle, the dilution concentration of the fatty acid ester is generally 0.2 wt% or more, preferably 0.2 to 10 wt%, and more preferably 0.5 to 5 wt%.

The solution containing the active principle is preferably prepared by diluting it as follows. The active principle such as the substance capable of controlling the cell membrane permeability of the hyphae (e.g., a surfactant and/or a natural vegetable oil) is dissolved in an organic solvent (preferably ethanol), and the resultant solution is stirred for 5 to 10 minutes to obtain a homogeneous solution. Thereafter, distilled water is added to the homogenous solution to obtain a solution containing the active principle at a desired concentration. The final concentration of the organic solvent is set at generally 0.2 to 5 wt%, preferably about 2 wt%. The organic solvent for use in preparing the solution is not particularly limited, as long as it does not negatively affect the growth of Matsutake hyphae. Ethanol is preferably used. For example, a solution containing 1 wt% Tween 80 can be prepared by mixing 1 g

of Tween 80 (commercially available) with 2 mL of 99.5% ethanol and then adding 98 mL of distilled water thereto. In this manner, it is possible to prepare a solution containing an active principle such as a surfactant and/or a natural vegetable oil homogenously (hereinafter also referred to as "a prepared solution containing an active principle").

(Preparation of culture substrate)

A culture substrate for adding the aforementioned active principle (such as a substance capable of controlling the cell membrane permeability of Matsutake hyphae) is not particularly limited, as long as Matsutake hyphae can grow therein. Both soil collected from a natural environment and an artificial substrate (for example, soil and vermiculite at a ratio of 1:1) may be used as the culture substrate. In Examples of the present invention, black soil was used, which contains a loamy layer of the Kanto Plain as a base.

Such a culture substrate is preferably sifted through a sieve to render the grain size of the substrate small and uniform. The grain size of the soil is adjusted preferably to about 3 mm or less, and more specifically about 2 to 3 mm. In the case of the artificial substrate, the grain size is preferably adjusted to about 2 mm or less, and more specifically about 1 to 2 mm.

The culture substrate that is made uniform in

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grain size as described above can be dried by dry heat. The dry heat is sufficient if it is performed at about 60°C for about 20-30 hours as an example.

To the dried culture substrate, the aforementioned
5 active principle is added. Preferably, to the dried culture substrate, the prepared solution containing an active principle such as a surfactant and/or a natural vegetable oil at a desired concentration, is added. At this time, the prepared solution is preferably
10 spread throughout the culture substrate such that the resultant culture substrate is not obtained in a wet state but a dry-touched state. In other words, the culture substrate is preferably prepared so as to contain the prepared solution in an amount of 15 to
15 30 wt% and more preferably 20 to 25 wt%, to the total weight. The active principle such as a surfactant and/or a natural vegetable oil is added to the culture substrate, and then it is sterilized and used as the culture substrate for Matsutake hyphae. The steriliza-
20 tion can be performed, for example, by autoclaving it at 120°C for 30 minutes or by γ -ray irradiation at 50-60 kGy.

The culture substrate prepared in this manner is suitable for growing Matsutake hyphae.

25 (Culture of Matsutake hyphae)

The culture substrate containing an active principle prepared as described above is put into

a sterile container (a Petri-dish, etc.). A hypha solution containing Matsutake hyphae is inoculated into the culture substrate in the sterile container such that the hypha solution is spread throughout the culture substrate. Culture of Matsutake hyphae can be performed at $25 \pm 1^\circ\text{C}$ in the dark. After 1 to 3 month's culture of Matsutake hyphae, a piece of the grown hyphae (i.e., a Shiro) are taken as a sample, and the growth amount of the hyphae is determined based on the amount of ergosterol. Ergosterol can be quantified as follows. After a sample of a Shiro is freeze-dried (EYELA, FDU-830), 1 g of the sample is taken and extracted with 2.5 mL of ethanol (99.5%). The obtained extract is loaded into HPLC (high-performance liquid chromatography; HITACHI, Column L-7300; UV Detector L-7400; Autosampler L-7200; Pump L-7100; Integrator D-7500) to quantify the amount of ergosterol in the extract. With respect to the details of the quantification procedure, reference should be made to Martin et al. (1990) Mycol. Res. 94: 1059-1064. About one week after the initiation of the culture, it is observed that the hyphae are grown on the surface of the culture substrate. About 4 weeks after the initiation of the culture, increase in the growth amount of the hyphae is found.

As described above, when Matsutake hyphae are cultured in a culture substrate containing an active

principle (e.g., a surfactant and/or a natural vegetable oil), the Matsutake hyphae grow with a remarkably larger growth amount than those cultured in a culture substrate containing no active principle.

5 According to the artificial Shiro formation method of the present invention, it is possible to form a Shiro containing hyphae in an amount corresponding to that of a natural Shiro in a Matsutake-producing forest, in a short culture period of several months. The artificial
10 Shiro formation method of the present invention is therefore extremely useful since it becomes a base for artificially cultivating Matsutake mushrooms.

(Preferred embodiments)

Preferred embodiments of the method of the present
15 invention will be explained below.

An artificial Shiro of Matsutake can be induced by culturing Matsutake hyphae in a culture substrate containing the aforementioned active principle such as a substance capable of controlling the cell membrane
20 permeability of Matsutake hyphae (e.g., a surfactant and/or a natural vegetable oil), as described above. Preferably, an inoculum of actively growing Matsutake hyphae is first prepared, and then the inoculum is cultured in a culture substrate containing the
25 aforementioned active principle. The method of preparing an inoculum of actively growing Matsutake hyphae comprises (1) a step of inducing growth

of matsutake hyphae and (2) a step of preparing an inoculum of matsutake hyphae.

These two steps will be explained below. Both of the steps are aseptically performed.

5 (1) Step of inducing growth of Matsutake hyphae

The step of inducing growth of Matsutake hyphae is performed by aseptically homogenizing a colony of Matsutake hyphae and aseptically culturing the obtained hyphae in a liquid nutrient medium.

10 In this step, as the colony of Matsutake hyphae, a colony of Matsutake hyphae which is aseptically subcultured on a solid medium (such as Ohta agar medium) may be used. Preferably, the colony (i.e., mycelia) may be prepared by taking a piece of the
15 colony subcultured on the solid medium, inoculating it in a liquid medium having the same composition, and further culturing it at 20-25°C in the dark for 2-3 weeks.

20 A piece of the aforementioned colony is taken out from the culture medium, a fresh liquid medium having the same composition is added to the colony, and it is homogenized aseptically, thereby obtaining a suspension of the hypha cells. The aseptic homogenization herein means that each hypha cell itself are not destroyed but
25 separated from each other. For example, the aseptic homogenization can be performed in a clean bench by using a dry heat sterilized homogenizer. The colony

may be homogenized with a homogenizer at about 80 × 100 rpm to 120 × 100 rpm, preferably 100 × 100 rpm, for about 4 to 8 seconds, preferably 6 seconds.

5 The suspension of the hypha cells that is obtained by the homogenization is further cultured for 3 to 5 days in the same liquid medium under the aforementioned conditions. In this manner, it is possible to obtain a suspension of actively growing Matsutake hyphae.

10 By homogenizing the colony as mentioned above, each of the hypha cells forming the colony is roughly separated from each other. By virtue of the separation, the Matsutake hypha cells having the capability to grow rapidly can be obtained. This is presumably because each of the separated hypha cells
15 intakes nutrition at a higher rate from the culture medium and it hastens the apical growth of the hypha cells, compared with the hypha cells in the form of a colony.

20 (2) Step of preparing an inoculum of Matsutake hyphae

The step of preparing an inoculum of Matsutake hyphae is performed by aseptically replacing the liquid nutrient medium containing the growth-induced Matsutake hyphae which is obtained in the aforementioned step
25 (1), with a liquid nutrient medium containing no carbon source.

More specifically, the preparation of the inoculum

can be performed as follows. The liquid nutrient medium containing Matsutake hyphae (preferably the liquid medium containing Matsutake hyphae having the capability to grow rapidly which is obtained in the
5 aforementioned step (1)) is first filtered, by use of a mesh such as a nylon mesh filter (average pore size $24 \times 30 \mu\text{m}$).

Next, the hyphae obtained by the filtration are aseptically rinsed with a liquid medium which
10 is modified so as not to contain a carbon source (hereinafter also referred to as "a modified liquid medium"). The rinsing is preferably performed at least three times with 10 mL of the modified liquid medium. The modified liquid medium for rinsing is not limited
15 as long as it is obtained by removing carbon source from a liquid medium capable of using for culturing the hyphae. Preferably, a modified liquid medium that is prepared by removing glucose from SH liquid medium is used (Schenk, R. U. & Hildebrandt, A. C. (1972) Can.
20 J. Bot. 50: 199-204).

After rinsing the hyphae, an appropriate amount of the modified liquid medium is added to the hyphae, thereby obtaining an inoculum of the hyphae wherein original liquid medium is replaced with the modified
25 liquid medium containing no carbon source.

When an artificial Shiro of Matsutake is induced using the inoculum of actively growing hyphae which is

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prepared by the steps (1) and (2), the advantageous effect of the present invention can be more remarkable.

(Examples)

5 The present invention will be explained in more detail by way of examples below, but the invention is not limited to these examples.

(Example 1)

Method of forming an artificial Shiro of Matsutake using a surfactant (Tween 80)

10 (1) Preparation of an inoculum of Matsutake hyphae

A piece of hyphae was taken from stock hyphae of a Matsutake strain (Tm 89; stored at the laboratory of the present inventors), and cultured on Ohta agar medium for 4 weeks. The composition of Ohta medium is shown below.

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Table 1 Ohta medium

Composition	Concentration (/1000 mL)
Glucose	10 g
Citric acid	1 g
Ammonium tartrate	1 g
KH ₂ PO ₄	1 g
MgSO ₄ ·7H ₂ O	1 g
CaCl ₂ ·2H ₂ O	50 mg
HEPES*	7 g
Mineral solution**	10 ml
Vitamin solution***	10 ml
pH	Adjusted to 5.1 with 1N KOH

* N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid

** Mineral solution(/1000 mL): FeCl₃ 5 mg,
MnSO₄·4-6H₂O 50 mg, ZnSO₄·7H₂O 300 mg, CoSO₄·7H₂O
50 mg, CuSO₄·5H₂O 100 mg, NiSO₄·6H₂O 200 mg,
Acetylacetone 3 mL

*** Vitamin solution(/1000 mL): Thiamin hydrochloride
300 mg, Nicotinic acid 5 mg, Folic acid 3 mg,
Biotin 5 mg, Pyridoxine hydrochloride 0.5 mg,
Carnitine chloride 1 mg, Adenine·H₂SO₄·2H₂O 3 mg,
Choline chloride 3 mg

The grown hyphae were cut in a size of about 2 mm
× 2 mm from the agar medium, and they were cultured
in fresh liquid Ohta medium for 3 weeks. A colony of
the obtained hyphae was homogenized with a homogenizer
and re-cultured for 3 days. After the culturing,
the resultant hyphae were rinsed three times with
a glucose-free liquid medium (a modified SH liquid
medium), and suspended in the same glucose-free liquid
medium. In this manner, an inoculum of Matsutake
hyphae was prepared. The composition of the
glucose-free medium is shown below.

Table 2 Modified SH liquid medium

Composition	Concentration (/1000 mL)
Ammonium tartrate	1000 mg
KH ₂ PO ₄	500 mg
MgSO ₄ ·7H ₂ O	200 mg
CaCl ₂ ·2H ₂ O	20 mg
FeSO ₄ ·7H ₂ O	15 mg
Na ₂ EDTA·H ₂ O	20 mg
H ₃ BO ₃	0.5 mg
ZnSO ₄ ·7H ₂ O	0.1 mg
MnSO ₄ ·H ₂ O	1 mg
Na ₂ MoO ₄ ·2H ₂ O	0.01 mg
KI	0.1 mg
CuSO ₄ ·5H ₂ O	0.02 mg
CoCl ₂ ·6H ₂ O	0.01 mg
Thiamin hydrochloride	0.5 mg
Nicotinic acid	0.5 mg
Pyridoxine hydrochloride	0.05 mg
pH	Adjusted to 5.1 with 1N NaOH

(2) Preparation of surfactant (Tween 80) additive
Tween 80 (Polyoxyethylene (20) Sorbitan
Monooleate, available from Wako Pure Chemical
Industries, Ltd.) was mixed with 99.5% ethanol and
5 stirred for 5 minutes. To the resultant mixture of
Tween 80/ethanol solution, distilled water was added to
prepare a Tween 80 additive. For example, 1 wt% of
Tween 80 additive was prepared by adding 1g of Tween 80
(commercially available) to 2 mL of 99.5% ethanol,
10 followed by adding 98 mL of distilled water. The final
concentration of ethanol resulted in about 2 wt%.

(3) Preparation of soil for culturing Matsutake
hyphae

Soil was taken from a site under a bush in the
15 campus of the University of Tokyo. The soil was sifted
through a sieve to adjust the grain size to 3 mm or
less. The soil (uniform in grain size) was dried at
60°C for 24 hours with a dry heat sterilizer. Tween 80
additive containing Tween 80 in amounts of 0 wt%,
20 1 wt%, 2 wt%, or 5 wt% (prepared in step (2) of Example
1) was added to the soil and mixed well. In the case
of the additive containing 0 wt% Tween 80, ethanol was
added in the same amount as the cases of the Tween 80
additives containing 1 wt%, 2 wt%, and 5 wt% Tween 80,
25 and ethanol concentration was 2% in each Tween 80
additive. The soil was prepared so as to contain
the Tween 80 additive in an amount of 20 wt% to the

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total weight, and then the soil was sterilized in an autoclave at 121°C for 30 minutes. In this manner, the soil for culturing Matsutake hyphae was prepared.

(4) Inoculation of Matsutake hyphae

5 The soil for culturing (prepared in step (3) of Example 1) was put in an amount of 30 mL to a sterile Petri-dish (90 mm diameter). 5 mL of Matsutake hyphae solution (i.e., the inoculum prepared in step (1) of Example 1) was inoculated into the soil in the Petri-dish so as to spread throughout the soil. Thereafter, 10 the Petri-dish was incubated at $25\pm 1^\circ\text{C}$ in the dark. One month after the initiation of the incubation, the amount of ergosterol, which is a component of the cell membrane of Matsutake hyphae, was measured to determine 15 the amount of the hyphae (Martin et al. (1990) Mycol. Res. 94: 1059-1064). The amount of ergosterol indicates the amount of Matsutake hyphae. The measurement of the amount of ergosterol was performed by collecting soil (1g) from each sample, extracting 20 ergosterol therefrom, and quantifying. As the control, soil was taken from a natural Shiro in a Matsutake-producing forest (Ina, Nagano prefecture, October, 2000) and subjected to the same measurement. From the measurement results of ergosterol (FIG. 1), it is found 25 that growth of Matsutake hyphae is significantly enhanced by the addition of Tween 80. On the other hand, the ergosterol amount in the natural Shiro

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derived from the Matsutake-producing forest (Ina) was
59.68 μ g/g dry soil.

(Example 2)

Method of forming an artificial Shiro of Matsutake
5 using natural vegetable oil (olive oil)

(1) Preparation of an inoculum of Matsutake
hyphae

Preparation of an inoculum of Matsutake hyphae
was performed in the same manner as described in the
10 step (1) of Example 1, except that Tm2 (stored in the
laboratory of the inventors) was used as the Matsutake
strain.

(2) Preparation of natural vegetable oil (olive
oil) additive

15 Olive oil (Ajinomoto Co. Inc.) was mixed with
99.5% ethanol, and stirred for 5 minutes. To the
resultant mixture of olive oil/ethanol solution,
distilled water was added to prepare an olive oil
additive. For example, an additive containing 1 wt%
20 olive oil was prepared by mixing 1g of olive oil
(commercially available) and 2 mL of 99.5% ethanol,
followed by adding 98 mL of distilled water. The final
concentration of ethanol resulted in about 2 wt%.

(3) Preparation of soil for culturing Matsutake
25 hyphae

Preparation of soil for culturing Matsutake hyphae
was performed in the same manner as described in the

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step (3) of Example 1, except that olive oil additive containing olive oil at amounts of 0%, 1%, or 2% (prepared in step (2) of Example 2) was added to the soil for culturing.

5 (4) Inoculation of Matsutake hyphae

 The soil for culturing (prepared in step (3) of Example 2) was put in an amount of 30 mL to a sterile Petri-dish (90 mm diameter). 5 mL of Matsutake hyphae solution (i.e., the inoculum prepared in step (1) of Example 2) was inoculated into the soil in the Petri-dish so as to spread throughout the soil. Thereafter, the Petri-dish was incubated at $25\pm 1^{\circ}\text{C}$ in the dark. Three months after the initiation of the incubation, the amount of ergosterol, which is a component of the cell membrane of the Matsutake hyphae, was measured to determine the amount of the hyphae (Martin et al. (1990) Mycol. Res. 94: 1059-1064). The measurement of the amount of ergosterol was performed by collecting soil (1g) from each sample, extracting ergosterol therefrom, and quantifying. As the control, soil was taken from a natural Shiro in a Matsutake-producing forest (Ina, Nagano prefecture, October, 2000) and subjected to the same measurement. From the measurement results of ergosterol (FIG. 2), it is found that the growth of Matsutake hyphae is significantly enhanced by the addition of olive oil. On the other hand, the amount of ergosterol in the natural Shiro

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(Advantages of the Invention)

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invention becomes a base for establishing a technique of artificial cultivation of Matsutake mushrooms.

Additional advantages and modifications will readily occur to those skilled in the art. Therefore, the invention in its broader aspects is not limited to the specific details and representative embodiments shown and described herein. Accordingly, various modifications may be made without departing from the spirit or scope of the general inventive concept as defined by the appended claims and their equivalents.